Cecropins Activity Against Bacterial Pathogens

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Abstract: Today, resistant bacteria are challenges in the treatment of bacterial infections. Cecropins are compounds that kill bacteria by affecting the membrane of microorganisms, such as Acinetobacter, Escherichia coli, and Staphylococcus. This study aimed to evaluate the antibacterial activity of cecropins against bacterial pathogens quantitatively. In this study, articles reporting antimicrobial activity of cecropins were searched in PubMed, Web of Science, and Scopus databases using the Google Scholar search engine. Then, the results of the current study were evaluated quantitatively. In this study, we found 29 studies reporting cecropins antimicrobial activity against major bacterial pathogens. Also, there were 25 studies on cecropin antimicrobial activity against gram-negative pathogens, and it was cleared that cecropin B antibacterial activity on Pseudomonas aeruginosa was lesser than others (minimum inhibitory concentration, 0.4 µg/ML), and we showed that Staphylococcus aureus growth can be inhibited by Cecropin AD more than others (minimum inhibitory concentration, 0.2 µg/Ml). Because cecropin peptides have no adverse effect on the human cells, and also, it has been demonstrated that cecropins have acceptable functions against pathogenic bacteria, we showed that they are potential candidates for research and construction of novel antibiotics.

Key Words: cecropins, peptides, antibacterial agents, resistance

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t present, antimicrobial resistance is rapidly expanding A throughout the world, so that the effectiveness of routine antibiotics has been questioned. Spreading antibiotic resistance and emergence of new resistant strains have created an urgent need to find a suitable alternative to antibiotics. In recent years, antimicrobial peptides (AMPs), which are components of the immune system, have been identified as effective antimicrobial agents for killing antibiotic-resistant bacteria. Antimicrobial peptides kill bacteria through 2 mechanisms. Antimicrobial peptide activity induces permeation in membranes of some bacteria, but in others, these compounds readily enter the cell membrane and interact with intracellular components, including nucleic acids.¹ An antimicrobial peptide is a cationic peptide and can affect gram-positive and gram-negative bacteria and mycobacterium growth.²⁻⁴ Cecropins are antibacterial peptides that were extracted originally from insects⁵; they consist of 31 to 37 amino acids, and can affect gram-negative and gram-positive bacteria and mycobacteria. Bactericidin, lepidoptin, and sarcotoxin are other names of these structurally related proteins.⁶ These peptides are secretory proteins and can be activated after deletion of the signal peptides, affecting a broad spectrum of gram-negative and gram-positive bacteria.⁷ Tuberculosis is another infectious disease with which many people, such as pregnant women⁸ throughout the world have been affected, and, in some cases, there is resistant to routine treatment mycobacterium. So, we searched antimycobacterial

Sciences, Esfarayen, Iran. E-mail: moradib901@gmail.com. The authors have no conflicts of interest to disclose. Copyright © 2020 Wolters Kluwer Health, Inc. All rights reserved. peptide killing Mycobacterium spp, and we found a few studies reporting antimycobacterial activity against Mycobacterium spp.^{9,10} Antiendotoxin activity is another property of cecropins that has been reported in a study.¹¹ The most important types of cecropins are cecropin A, cecropin B, and cecropin P1. So far, a few studies have been performed on the antibacterial effect of cecropins on bacterial pathogens.^{12,13} Cecropin A includes the KWKLFKKIEKVGQNIRDGIIKAGPAVAVVGQATQIAK amino acid sequence, in which the secondary structure consists of 2 α helices.² Cecropin A, like other cecropins can affect the microbial cytoplasmic membrane, and this property has been demonstrated. In one study, antibacterial activity of cecropins was determined against multidrug-resistant nosocomial isolates of Acinetobacter baumannii compared with melittin and cecropin A-melittin peptide CA.14 Of course, it is necessary to mention all kinds of cecropins can damage microbial membrane by 1 mechanism, and they can affect both gram-positive and gram-negative pathogens. Cecropin B amino acid have been sequenced (KWKVFKKIEKMGRNIRNGIVKAGPAIAVLGEAKAL) and structured by 2 α helices.² Cecropin B was extracted from the giant silkmoth, Hyalophora cecropia, by researchers and its antibacterial activity have been determined. This peptide is belonging to innate immune system and is effective protein in clearance and promotion of wound repair.1

Furthermore, the antibacterial activity of this peptide has been evaluated against both gram-negative and gram-positive bacteria. ^{16,17} The antibacterial activity of cecropin B also is related to the interaction with bacterial LPS so that disruption of LPS causes killing of bacteria. Cecropin B also inhibit production of cytokines nitric oxide and release of tumor necrosis factor- α in the host. So, these reports confirm that cecropin B have a potential in antibacterial and anti-inflammatory activities.

Finally, cecropin P1 (CP1) is a 3.3-kDa, 31-amino acid, that amino acid sequence contains SWLSKTAKKLENSAKKRISEGI AIAIQGGPR. This antibacterial peptide has been isolated from Ascaris, an intestinal nematode of pigs.¹ In one study, antibacterial mechanism of cecropin P1 have been shown, in which cecropin P1 can obstruct bacterial membranes instantaneously, and also, it was cleared that CP1 antimicrobial activity is time- and dose-dependent and but CP1 permeability begin at low concentrations.¹⁸ In another study, there is evidences in which CP1 can be active against important infectious bacteria, such as *Pseudomonas aeruginosa*¹⁹ and antibacterial resistance for cecropin P1 could not be considered because CP1 action on bacterial membrane is performed physically. Here we can mention a property of CP1 and that is lytic action without hemolysis.²⁰

Although the antimicrobial profile of many antibiotics^{21–23} and other antimicrobial agents²⁴ in exposure to bacterial pathogens have been determined so far, no comprehensive efforts have been made for AMPs till now. Therefore, the aim of our study was reporting in vitro quantitative activity of cecropin A, cecropin B, and cecropin P1 against multidrug-resistant bacteria.

MATERIAL AND METHODS

In the present study, we searched articles in Chemical Abstract, EBSCO, Scopus, PubMed, ISC, Web of Science, and

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			A			
Gram Negative Bacteria	Strain	Cecropin	(MIC µg/mL)	(MBC µg/mL)	Other (µg/mL)	References
P. aeruginosa	PA14	Cecropin A	32	ND	ND	25
	PA103		64	ND	ND	
	PA 2326		64	ND	ND	
	PA 1026		32	ND	ND	
	PAO1		64	ND	ND	
	PA 1016		>64	ND	ND	
	OT97		ND	ND	2.6	26
		Cecropin B	ND	ND	1.5	
	PAO1	CEME	2	ND	ND	27
		CE	>64	ND	ND	28
	ATCC 27853	BP100	33.5	ND	ND	29
		Cecropin P1	>256	ND	ND	30
		Cecropin B	64	ND	ND	
		CE	>64	ND	ND	28
	Clinical	Cecropin P1	4–64	8-128	ND	31
	ATCC27853	Cecropin B	0.40	ND	ND	32
		Cecropin P1	0.8-1.60	ND	ND	
Pseudomonas	Pseudomonas fluorescent	Cecropin 1	1.1	ND	ND	33
	·	Cecropin 2	1.9	ND	ND	
		Cecropin 3	1.3	ND	ND	
K. pneumoniae	ATCC77326	Cecropin A	4	ND	ND	25
1	ATCC 700603 (MDR)	BP100	17	ND	ND	29
	Clinical strain	Cecropin P1	0.25-2	0.50-4	ND	31
Escherichia coli	OP50	Cecropin A	4	ND	ND	34
	WT	- · · · I	0.5	ND	ND	
	WT GFP		0.5	ND	ND	
	Δ waaP GFP		0.5	ND	ND	
	Δ waaC GFP		0.5	ND	ND	
	Δ waa F		0.25	ND	ND	
	Δ waa I		0.25	ND	ND	
	Δ waa Y		0.25	ND	ND	
	MG1655		0.9	ND	ND	
	D31		64-78	ND	ND	21
	Clinical		2.5	ND	ND	35.36
		Cecropin B	2.5	ND	ND	,
		Cecropin pl	0.1-4	ND	ND	
	DH5 a	DAN1	4.9	ND	ND	37
	Diffe d	DAN2	2.1	ND	ND	57
	Standard	Cecropin A	0.8-5	ND	ND	38
	D21	e veropii i i	ND	ND	0.4	26
	D21	Cecronin B	ND	ND	0.4	20
	SC9251	Cecropin B	2	ND	ND	39
	UB1005	CEME	- 1	ND	ND	27
	D31	Cecropin A	1	ND	ND	40
	G	Cecropin A B	0.1	ND	ND	41
	K12	Cecropin-like protein	0.15	ND	ND	42
	Clinical	Cecropin A	3	20	ND	43
	DH50	Cecronin 1	0.1	20 ND	ND	-1.5
	DIDU	Cecropin 2	0.1	ND	ND	55
		Cecropin 2	0.5			
	ATCC 25022	Cecropin A	64			26
	Coli LIP 100	Cecropin A	04 \\61			20
	COILUD 100	Cectopiii A	~04	ND	IND	

TABLE 1. Quantitative Antimicrobial Activity of Cecropins Against Gram-negative Bacteria

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TABLE 1. (Continued)

			Antibacterial Activity			
Gram Negative Bacteria	Strain	Cecropin	(MIC µg/mL)	(MBC µg/mL)	Other (µg/mL)	References
	ER2566	Cecropin B2	6.8	ND	ND	44
	BL21		7.2	ND	ND	
	Rosetta		7.2	ND	ND	
	JM109		6.6	ND	ND	
	DH1		6.7	ND	ND	
	K12D31	Cecropin AD	1.8	ND	ND	45
	K88		2	ND	ND	
	K99		2	ND	ND	
	UB100	CEME	2	ND	ND	46
	ATCC25922	Cecropin P1	16-32	ND	ND	30
		Cecropin B	16-32	ND	ND	
		BP100	17	ND	ND	29
	Clinical strain	Cecropin P1	0.25-1	0.25-2	ND	31
		Cecropin B	16	ND	ND	47
	ATCC25922	Cecropin B	0.4-1.60	ND	ND	32
		Cecropin P1	0.8-1.60	ND	ND	
	HB10	Cecropin B	0.10	ND	ND	
		Cecropin P1	0.8-1.60	ND	ND	
Acinetobacter baumannii	ATCC 17978	Cecropin A	2	ND	ND	25
	ATCC19606	-	4	8	ND	48
	GIM1.650		4	8	ND	
	Ac157	Cecropin A-Melittin	2	ND	ND	49
	BCRC 15884	Cecropin B2	6.8	ND	ND	44
	E1359		6.9	ND	ND	
	ATCC BAA-1605 (MDR)	Cecropin- α -melittin	17-67	ND	ND	29
	Colistin susceptible	(BP100)	8.5-18	ND	ND	
Acinetobacter spp	Clinical strain	Cecropin P1	0.50-2	1–4	ND	31
		Cecropin B	16	ND	ND	47
Shigella sonnei	JS1 1746	Cecropin-like protein	0.08	ND	ND	42
Shigella spp.	clinical	Cecropin P1	1-32	2-32	ND	
Proteus vulgaris	OX19	Cecropin-like protein	0.3	ND	ND	
S. typhimurium	ATCC 14028	CE	>64	ND	ND	28
	ATCC 7731		>64	ND	ND	
	Clinical strain	Cecropin AD	8	ND	ND	45
Salmonella typhi	Clinical strain	Cecropin P1	0.50-8	1-8	ND	
S. enteritidis	Clinical strain	cecropin AD	16	ND	ND	
S. entericaser. Typhimurium	L T2	Cecropin P1	>128	ND	ND	30
<i>v</i> 1		Cecropin B	32	ND	ND	
Brucella spp	Clinical strain	Cecropin P1	0.25-2	0.50-2	ND	31
Francisella spp	F. novicida	Cecropin A1	ND	ND	20.1	22
**		Cecropin B	ND	ND	4.64	
ND. not determined.						

MedLib databases. Our search strategy was included: ("Cecropin "(All Fields)) AND (peptide"(All Fields)) AND resistance (All Fields) AND Bacteria (All Fields) AND Antimicrobial (All Fields) and keywords selected from Medical Subject Headings thesaurus. Articles were searched online and without time limitation, and inclusion criteria for articles in this study were based on report of quantitative evaluation of minimum inhibitory concentration minimum inhibitory concentration (MIC), minimum bactericidal concentration (MBC), cecropins, and bacterial pathogens. In contrast, exclusion criteria included inability to access the full text and reports containing unrelated results. Data analysis in this review study was performed based on the reports, and data analysis methods or tools such as SPSS were not used.

RESULTS

In this study we found 71 articles which had reported the antimicrobial profile of cecropins and derivatives quantitatively. After examination of related articles, it was revealed that 29 studies had quantitative reports of antimicrobial activity of cecropins against bacterial pathogens based on MIC, MBC, and lethal dose. There were 25 studies on cecropin antimicrobial activity against

Gram-positive Bacteria	Strains	Cecropin	(MIC µg/mL)	(MBC µg/mL)	Other (µg/mL)	References
Staphylococcus aureus	RN4220	CEME	4		47	
1 2	MW2	Cecropin A2	>64	ND	ND	25
	MRSA ATCC43300	CAMA	8	ND	640	50
	Standard	Cecropin 1	2.1	ND	ND	33
		Cecropin 2	5	ND	ND	
		Cecropin 3	2.5	ND	ND	
	ATCC29213	DAN1	4.9	ND	ND	37
		DAN2	2.1	ND	ND	
		Cecropin A	>64	ND	ND	28
		Cecropin P1	>256	ND	ND	30
		Cecropin B	>256	ND	ND	
	MRSA	Cecropin P1	>128	>128	ND	31
	MS	Cecropin P1	32128	32128	ND	
	MR	Cecropin P1	8-128	16-128	ND	
	IVDC C56005	Cecropin AD	0.2	ND	ND	45
	25923	CEME	8	ND	ND	46
	SAP0017 (MRSA)		4	ND	ND	
	Clinical isolate		8	ND	ND	
	Clinical isolate		4	ND	ND	
	ATCC 33591 (MRSA)	Cecropin-α-melittin hybridBP100	134.5	ND	ND	29
	ATCC9144	Cecropin P1	>100	ND	ND	32
	clinically resistant	Cecropin B	25	ND	ND	
	·		32	ND	ND	47
Streptococcus faecalis	IVDC C55614	Cecropin AD	24	ND	ND	45
Enterococcus faecium	E007	Cecropin A2	>64	ND	ND	25
-	ATCC 700221 (VRE)	Cecropin-α-melittin hybridBP100	67	ND	ND	29
Enterococcus faecalis	ATCC 29212	Cecropin A	>64	ND	ND	28
		CEME	32	ND	ND	46
		Cecropin P1	>256	ND	ND	30
		Cecropin B	>256	ND	ND	
	Clinical isolate	Cecropin P1	>128	>128	ND	31
S. epidermidis	ATCC12228	Cecropin A	>64	ND	ND	28
	Clinical isolate	CEME	8	ND	ND	46
S. pyogenes	ATCC 19615		8	ND	ND	
Listeria. monocytogenes	NCTC 7973		4	ND	ND	
	N22-2	Cecropin P1	>256	ND	ND	30
		Cecropin B	>256	ND	ND	
Streptococcus pneumoniae	Clinical isolate	Cecropin P1	8-128	8-128	ND	31

TABLE 2. Quantitative Antimicrobial Activity of Cecropins Against Gram-positive Bacteria

gram-negative pathogens and it was cleared that cecropin B antibacterial activity on *P. aeruginosa* was lesser than others (MIC, 0.4 μ g/mL) (Table 1), 11 studies against gram-positive pathogens and we found *Staphylococcus aureus* growth can be inhibited by Cecropin AD more than others (MIC, 0.2 μ g/mL) (Table 2), and 3 studies against mycobacterial pathogens (Table 3).

DISCUSSION

Cecropins are known antimicrobial cationic peptides that can inhibit the growth and activity of bacterial pathogens. These cationic peptides were extracted originally from insects,²¹ but later, it was revealed that cecropins are a part of mammalian host innate immunity and probably can affect pathogenicity of microbial infection.^{22–24} These results are according to our study findings as our results showed cecropins are antibacterial peptides that can inhibit important strains of pathogenic bacteria. Mechanisms of this antibacterial AMP have been determined first in Christensen et al⁵¹ study, and it seems a strong positive charge by cecropins can form specific channels in the bacterial membrane by negative charge. It is necessary to mention that accordance to⁵² Li et al study, cecropins are not toxigenic for human cell membranes due to lower negative charge in mammalian cell membranes. And in the Schweizer study,⁵³ it has been revealed that cecropin B have selective toxicity. So, if appropriate

Strains	Cecropin	(MIC µg/mL)	(MBC µg/mL)	Other (µg/mL)	References
Mtb H37Rv	Cecropin P1	50,000	ND	ND	9
Mtb H37Ra	Cecropin B	600	ND	ND	10
CI74		1200	ND	ND	
CI85		1200	ND	ND	
CI114		600	ND	ND	
ClI121		1200	ND	ND	
CI		>120	ND	ND	4
	Cecropin A	>120	ND	ND	
CI, clinical isola	ite.				

	TABLE 3.	Quantitative	Antimicrobial	Activity of	Cecropins	Against <i>M.</i>	tuberculosis
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concentration of cecropins is adjacent to the bacterial membrane, the membrane will be altered due to difference in charge, and channels will be formed that disrupt the osmotic balance of the bacteria.⁵⁴ So high concentration of some cecropin such as cecropin B reported in the present study against important bacteria, such as *Mycobacterium tuberculosis*, could be considered as an antibiotic candidate (MIC>120).

Also, in the present study we reported the antimicrobial effects of cecropin based peptides on human pathogenic gram-negative bacteria, that is according to Moore et al study.¹² We evaluated cecropins activity on gram negative bacteria and it was found that cecropins can inhibit gram negative bacteria growth that have been mentioned in Table 1. For example, it was cleared that cecropin B antibacterial activity on P. aeruginosa was 0.4 µg/mL. After exanimation of evaluated articles, we found that some cecropins had antimicrobial activity also against gram-positive bacteria, ac-cording to Moore et al Wang et al^{12,55} studies, that have been mentioned in Table 2. In the present study result showed that gram positive pathogens such as Staphylococcus aureus growth can be inhibited by 0.2 µg/mL of Cecropin AD. Furthermore, our data showed that cecropins B, P1 can inhibit Mycobacterium genus growth in high concentration of cecropins. This result is according to Siemion et al,⁵⁶ Linde et al,⁹ and Portell-Buj et al⁴ examination results. Our study showed that some cecropin peptides such as cecropin AD (MIC = $0.2 \mu g/Ml$) and cecropin P1 (MIC = $0.25-2 \mu g/Ml$) could be considered a safe antibacterial agent for human infection treatments in a safe concentration. We found that gram-negative bacteria, such as Escherichia coli, P. aeruginosa, and Acinetobacter baumannii, can be killed by cecropins. Also, it was revealed that cecropins had antibacterial property against Enterococci, Staphylococcus aureus, and Streptococcus pyogenes. Effective antibacterial activity of cecropins depends on several factors, such as the kind of bacterial strain and standard dosage of cecropins. Some strains, such as multidrug-resistant A. baumannii and K. pneumoniae, were inhibited by the high dosage of cecropins, but in the low dosage only susceptible strains had been inhibited. But, in resistant gram-positive bacteria, such as methicillin-resistant S. aureus and vancomycin-resistant Enterococcus, cecropin EME, and BP100 had opposite properties. Inhibition of susceptible strains by lower dosage of cecropins is in accordance with another study performed on AMP.57 Tuberculosis is another infectious disease by which many people throughout the world have been affected, and in some cases, there is resistant to routine treatment mycobacterium. Therefore, we searched antimycobacterial peptide killing *Mycobacterium* spp., and we found a few studies reporting antimycobacterial activity against *Mycobacterium* spp.^{9,10} After examination of more searched articles, we found that there were no further similar studies conducted on quantitative evaluation of cecropin antibacterial profile. However, we found a few articles reporting in vitro antimicrobial activity of cecropins on bacterial pathogens^{58–61} and on antimicrobial activity of AMPs against viruses and fungi.⁶⁰

Studies on cationic peptides have increased recently. Because these proteins have no adverse effects on human cells and have also shown an acceptable function against pathogenic bacteria, they are known as potential antibiotic candidates. Today, a number of these proteins, such as cecropin based peptides D2A21 and D4E1 have been entered into clinical trials assessment.^{62,63} Therefore, it is proposed that other kinds of cecropin peptides be evaluated in vitro and in vivo and in animal models.

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REFERENCES

- Mookherjee N, Hancock R. Cationic host defence peptides: innate immune regulatory peptides as a novel approach for treating infections. *Cell Mol Life Sci.* 2007;64:922–933.
- Hoskin DW, Ramamoorthy A. Studies on anticancer activities of antimicrobial peptides. *Biochim Biophys Acta*. 2008;1778:357–375.
- Gutsmann T. Interaction between antimicrobial peptides and mycobacteria. Biochim Biophys Acta. 1858;2016:1034–1043.
- Portell-Buj E, Vergara A, Alejo I, et al. In vitro activity of 12 antimicrobial peptides against Mycobacterium tuberculosis and Mycobacterium avium clinical isolates. J Med Microbiol. 2019;68(2):211–215.
- Gazit E, Boman A, Boman HG, et al. Interaction of the mammalian antibacterial peptide cecropin P1 with phospholipid vesicles. *Biochemistry*. 1995;34(36):11479–11488.
- Boman HG, Faye I, Gudmundsson GH, et al. Cell-free immunity in Cecropia. A model system for antibacterial proteins. *Eur J Biochem.* 1991; 201(1):23–31.
- Cavallarin L, Andreu D, San Segundo B. Cecropin A-derived peptides are potent inhibitors of fungal plant pathogens. *Mol Plant-Microbe Interact*. 1998;11:218–227.
- Moradi B, Meshkat Z. Evaluation of tuberculosis infection in pregnant women and its effects on newborns: an overview. *IJOGI*. 2015;18:21–36.
- Linde CM, Hoffner SE, Refai E, et al. In vitro activity of PR-39, a proline-arginine-rich peptide, against susceptible and multi-drug-resistant *Mycobacterium tuberculosis. J Antimicrob Chemother.* 2001;47(5): 575–580.

- Santos P, Gordillo A, Osses L, et al. Effect of antimicrobial peptides on ATPase activity and proton pumping in plasma membrane vesicles obtained from mycobacteria. *Peptides*. 2012;36:121–128.
- Scott MG, Rosenberger CM, Gold MR, et al. An alpha-helical cationic antimicrobial peptide selectively modulates macrophage responses to lipopolysaccharide and directly alters macrophage gene expression. *J Immunol.* 2000;165:3358–3365.
- Moore AJ, Beazley WD, Bibby MC, et al. Antimicrobial activity of cecropins. J Antimicrob Chemother. 1996;37(6):1077–1089.
- Fitriyanti M, Narsimhan G. Synergistic effect of low power ultrasonication on antimicrobial activity of cecropin P1 against E. coli in food syste. *LWT*. 2018;96:175–181.
- Giacometti A, Cirioni O, Kamysz W, et al. Comparative activities of cecropin A, melittin, and cecropin A-melittin peptide CA(1-7)M(2-9)NH2 against multidrug-resistant nosocomial isolates of *Acinetobacter baumannii*. *Peptides*. 2003;24(9):1315–1318.
- Kulagina NV, Shaffer KM, Ligler FS, et al. Antimicrobial peptides as new recognition molecules for screening challenging species. *Sensors Actuators B Chem.* 2007;121(1):150–157.
- Lee PH, Rudisill JA, Lin KH, et al. HB-107, a nonbacteriostatic fragment of the antimicrobial peptide cecropin B, accelerates murine wound repair. *Wound Repair Regen*. 2004;12(3):351–358.
- Bland JM, De Lucca AJ, Jacks TJ, et al. All-D-cecropin B. synthesis, conformation, lipopolysaccharide binding, and antibacterial activity. *Mol Cell Biochem*. 2001;218:105–111.
- Boman HG, Agerberth B, Boman A. Mechanisms of action on *Escherichia coli* of cecropin P1 and PR-39, two antibacterial peptides from pig intestine. *Infect Immun.* 1993;61(7):2978–2984.
- Giacometti A, Cirioni O, Barchiesi F, et al. In-vitro activity of cationic peptides alone and in combination with clinically used antimicrobial agents against *Pseudomonas aeruginosa*. J Antimicrob Chemother. 1999;44:641–645.
- Vunnam S, Juvvadi P, Merrifield R. Synthesis and antibacterial action of cecropin and proline-arginine-rich peptides from pig intestine. *J Pept Res.* 1997;49:59–66.
- Chernysh S, Gordya N, Suborova T. Insect antimicrobial peptide complexes prevent resistance development in bacteria. *PLoS One.* 2015; 10:e0130788.
- Kaushal A, Gupta K, Shah R, et al. Antimicrobial activity of mosquito cecropin peptides against Francisella. *Dev Comp Immunol.* 2016;63: 171–180.
- Dutta P, Das S. Mammalian antimicrobial peptides: promising therapeutic targets against infection and chronic inflammation. *Curr Top Med Chem.* 2016;16:99–129.
- Lee JY, Boman A, Sun CX, et al. Antibacterial peptides from pig intestine: isolation of a mammalian cecropin. *Proc Natl Acad Sci U S A*. 1989;86(23): 9159–9162.
- Zheng Z, Tharmalingam N, Liu Q, et al. Synergistic efficacy of *Aedes* aegypti antimicrobial peptide cecropin A2 and tetracycline against *Pseudomonas aeruginosa. Antimicrob Agents Chemother.* 2017;61(7): e00686–e00617.
- Li ZQ, Merrifield RB, Boman IA, et al. Effects on electrophoretic mobility and antibacterial spectrum of removal of two residues from synthetic sarcotoxin IA and addition of the same residues to cecropin B. *FEBS Lett.* 1988;231:299–302.
- Friedrich C, Scott MG, Karunaratne N, et al. Salt-resistant alpha-helical cationic antimicrobial peptides. *Antimicrob Agents Chemother*. 1999;43: 1542–1548.
- Tan T, Wu D, Li W, et al. High specific selectivity and membrane-active mechanism of synthetic cationic hybrid antimicrobial peptides based on the peptide FV7. *Int J Mol Sci.* 2017;18(2):339.
- Oddo A, Thomsen TT, Kjelstrup S, et al. An amphipathic undecapeptide with all d-amino acids shows promising activity against colistin-resistant

strains of acinetobacter baumannii and a dual mode of action. *Antimicrob Agents Chemother*. 2016;60(1):592–599.

- Ebbensgaard A, Mordhorst H, Overgaard MT, et al. Comparative evaluation of the antimicrobial activity of different antimicrobial peptides against a range of pathogenic bacteria. *PLoS One.* 2015;10:e0144611.
- Giacometti A, Cirioni O, Greganti G, et al. In vitro activities of membrane-active peptides against gram-positive and gram-negative aerobic bacteria. *Antimicrob Agents Chemother*. 1998;42:3320–3324.
- Paulsen VS, Blencke H-M, Benincasa M, et al. Structure-activity relationships of the antimicrobial peptide arasin 1—and mode of action studies of the N-terminal, proline-rich region. *PLoS One.* 2013;8:e53326.
- Ouyang L, Xu X, Freed S, et al. Cecropins from *Plutella xylostella* and their interaction with *Metarhizium anisopliae*. *PLoS One*. 2015;10:e0142451.
- 34. Agrawal A, Weisshaar JC. Effects of alterations of the *E. coli* lipopolysaccharide layer on membrane permeabilization events induced by cecropin A. *Biochim Biophys Acta Biomembr.* 1860;2018:1470–1479.
- Miskimins Mills BE. Modulatory activities of glycosaminoglycans and other polyanionic polysaccharides on cationic antimicrobial peptides. 2010.
- Giacometti A, Cirioni O, Barchiesi F, et al. In vitro susceptibility tests for cationic peptides: comparison of broth microdilution methods for bacteria that grow aerobically. *Antimicrob Agents Chemother*. 2000;44:1694–1696.
- Duwadi D, Shrestha A, Yilma B, et al. Identification and screening of potent antimicrobial peptides in arthropod genomes. *Peptides*. 2018;103: 26–30.
- Pöppel A-K, Vogel H, Wiesner J, et al. Antimicrobial peptides expressed in medicinal maggots of the blow fly *Lucilia sericata* show combinatorial activity against bacteria. *Antimicrob Agents Chemother*. 2015;59: 2508–2514.
- Vaara M, Vaara T. Ability of cecropin B to penetrate the enterobacterial outer membrane. *Antimicrob Agents Chemother*. 1994;38:2498–2501.
- Rahnamaeian M, Cytryńska M, Zdybicka-Barabas A, et al. The functional interaction between abaecin and pore-forming peptides indicates a general mechanism of antibacterial potentiation. *Peptides*. 2016;78:17–23.
- Oh J-T, Cajal Y, Skowronska EM, et al. Cationic peptide antimicrobials induce selective transcription of micF and osmY in *Escherichia coli*. *Biochim Biophys Acta*. 2000;1463:43–54.
- 42. Okada M, Natori S. Purification and characterization of an antibacterial protein from haemolymph of *Sarcophaga peregrina* (flesh-fly) larvae. *Biochem J.* 1983;211:727–734.
- Fieck A, Hurwitz I, Kang AS, et al. *Trypanosoma cruzi*: synergistic cytotoxicity of multiple amphipathic anti-microbial peptides to *T. cruzi* and potential bacterial hosts. *Exp Parasitol.* 2010;125(4):342–347.
- Lai WS, Kan SC, Lin CC, et al. Antibacterial peptide cecropinB2 production via various host and construct syste. *Molecules*. 2016;21:103.
- Chen X, Zhu F, Cao Y, et al. Novel expression vector for secretion of cecropin AD in *Bacillus subtilis* with enhanced antimicrobial activity. *Antimicrob Agents Chemother*. 2009;53:3683–3689.
- Scott MG, Gold MR, Hancock RE. Interaction of cationic peptides with lipoteichoic acid and gram-positive bacteria. *Infect Immun.* 1999;67(12): 6445–6453.
- Joshi S, Bisht GS, Rawat DS, et al. Comparative mode of action of novel hybrid peptide CS-1a and its rearranged amphipathic analogue CS-2a. *FEBS J.* 2012;279:3776–3790.
- Gui S, Li R, Feng Y, et al. Transmission electron microscopic morphological study and flow cytometric viability assessment of *Acinetobacter baumannii* susceptible to *Musca domestica cecropin*. *ScientificWorldJournal*. 2014;2014:657536.
- Saugar JM, Alarcón T, López-Hernández S, et al. Activities of polymyxin B and cecropin A-melittin peptide CA(1-8)M(1-18) against a multiresistant strain of *Acinetobacter baumannii*. *Antimicrob Agents Chemother*. 2002; 46:875–878.

- Mataraci E, Dosler S. In vitro activities of antibiotics and antimicrobial cationic peptides alone and in combination against methicillin-resistant *Staphylococcus aureus* biofilms. *Antimicrob Agents Chemother*. 2012;56: 6366–6371.
- Christensen B, Fink J, Merrifield RB, et al. Channel-forming properties of cecropins and related model compounds incorporated into planar lipid membranes. *Proc Natl Acad Sci U S A*. 1988;85(14):5072–5076.
- Li J, Koh JJ, Liu S, et al. Membrane active antimicrobial peptides: translating mechanistic insights to design. *Front Neurosci*. 2017;11:73.
- Tamang DG, Saier MH Jr. The cecropin superfamily of toxic peptides. J Mol Microbiol Biotechnol. 2006;11(1-2):94–103.
- Abrunhosa F, Faria S, Gomes P, et al. Interaction and lipid-induced conformation of two cecropin-melittin hybrid peptides depend on peptide and membrane composition. *J Phys Chem.* 2005;109:17311–17319.
- Wang YZ, Xu ZR. Comparison of antimicrobial activity in vitro of antimicrobial peptides and antibiotics against gram-positive and gram-negative bacteria. *Chin J Vet Sci.* 2004;24(3): 270–273.
- Siemion I, Gawłowska M, Wieczorek Z. Mycobacterium kansasii phagocytosis inhibition by the oligopeptides derived from systemine, cecropin a and BRCT-1 protein sequences. Pol J Chem. 2004;78: 1073–1079.

- Dosler S, Gerceker AA. In vitro activities of antimicrobial cationic peptides; melittin and nisin, alone or in combination with antibiotics against gram-positive bacteria. J Chemother. 2012;24(3):137–143.
- Sathyamoorthi A, Kumaresan V, Palanisamy R, et al. Therapeutic cationic antimicrobial peptide (CAP) derived from fish aspartic proteinase cathepsin D and its antimicrobial mechanism. *Int J Pept ResTher*. 2019;25: 93–105.
- Lu L, Huang X, Zhang L, et al. Antimicrobial peptides characteristics and their application in domestic animals. *J Anim Vet Adv.* 2014;13:1185–1193.
- Liu X, Guo C, Huang Y, et al. Inhibition of porcine reproductive and respiratory syndrome virus by cecropin D in vitro. *Infect Genet Evol*. 2015; 34:7–16.
- Montesinos L, Bundó M, Izquierdo E, et al. Production of biologically active cecropin a peptide in rice seed oil bodies. *PLoS One*. 2016; 11:e0146919.
- Cummins JE Jr., Guarner J, Flowers L, et al. Preclinical testing of candidate topical microbicides for anti-human immunodeficiency virus type 1 activity and tissue toxicity in a human cervical explant culture. *Antimicrob Agents Chemother*. 2007;51(5):1770–1779.
- Gondhali B, More T, Naik R, et al. In vitro bioefficacy of different antimicrobial peptides against different pathovers of xanthomonas using paper disc and micro-dilution broth method. *J Pure ApplMicrobiol*. 2016; 10:1251–1261.